

Chan et al.:

# Bottom-Up Physiologically-Based Biokinetic Modelling as an Alternative to Animal Testing

## Supplementary Data

### Results S1: Derivation of Inputs into Mech KiM model for Pitavastatin

In the kidney, pitavastatin is a substrate of OAT3 with  $K_m = 3.3 \mu\text{M}$  (Fujino et al., 2005). To obtain  $J_{max}$ ,  $CL_{int}$  from experiments conducted in kidney slices were used (Watanabe et al., 2011).

$$CL_{int} = \frac{(2.55 + 4.36)}{2} \text{ ml/g kidney/15 minutes} \div 15 \text{ minutes} \div (60 \times 10^6) \text{ PTC/g kidney}$$
$$= 3.84 \mu\text{l/min}/10^6 \text{ PTC}$$

$$J_{max} = CL_{int} \times K_m$$
$$= 3.84 \mu\text{l/min}/10^6 \text{ PTC} \times 3.3 \mu\text{M}$$
$$= 12.672 \text{ pmol/min}/10^6 \text{ PTC}$$

### Results S2: Derivation of Inputs for OATP1B3 and OATP2B1 Transport of Rosuvastatin

All values used are taken from Bosgra et al. (2014).

1. Determine the average expression of total amount of protein per million HEK-293 (Y):

$$CL_{int} (\text{l/h}) = \text{in vitro } J_{max} (\text{pmol/min/mg protein}) \times Y (\text{mg protein}/10^6 \text{ HEK}) \times \text{REF} \times \text{HPPGL} \times \text{liver weight (g)} \div K_m (\mu\text{M})$$

$$Y_{\text{OATP1B1}} = \frac{241 \times 13}{202 \times \frac{25.7}{9.8} \times 139 \times 1561} \div 60 \text{ min/h} \times 10^6 = 0.4543 \text{ mg protein}/10^6 \text{ HEK cells}$$

$$Y_{\text{OATP1B3}} = \frac{145 \times 16.5}{251 \times \frac{2.2}{1.4} \times 139 \times 1561} \div 60 \text{ min/h} \times 10^6 = 0.4659 \text{ mg protein}/10^6 \text{ HEK cells}$$

$$Y_{\text{OATP2B1}} = \frac{649 \times 26.1}{26.2 \times \frac{24.2}{0.2} \times 139 \times 1561} \div 60 \text{ min/h} \times 10^6 = 0.4104 \text{ mg protein}/10^6 \text{ HEK cells}$$

$$\text{Average Y} = \frac{0.4543 + 0.4659 + 0.4104}{3} = 0.4435 \text{ mg protein}/10^6 \text{ HEK cells}$$

2.  $J_{max}$  of OATP1B3 in HEK-293 cells

$$J_{max} = 251 \text{ pmol/min/mg protein} \times 0.4435 \text{ mg protein}/10^6 \text{ HEK}$$
$$= 111.3 \text{ pmol/min}/10^6 \text{ HEK cells}$$

3.  $CL_{int}$  of OATP2B1 in HEK-293 cells

$$J_{max} = 26.2 \text{ pmol/min/mg protein} \times 0.4435 \text{ mg protein}/10^6 \text{ HEK}$$
$$= 11.62 \text{ pmol/min}/10^6 \text{ HEK cells}$$

As the REF value for OATP2B1 (148.830) was greater than the allowed input value of 100,  $J_{max}$  was scaled up by 1.4883 times,

$$J_{max} = 11.62 \times 1.4883$$
$$= 17.3 \text{ pmol/min}/10^6 \text{ HEK cells}$$

### Results S3: Derivation of Inputs for Intestinal OATP2B1 Transport of Pitavastatin

From Equation 11:

$$P_{app,trans,n,i} (10^{-6} cm/s) = \frac{J_{max} (pmol/min)}{A (cm^2) \times [(K_m (\mu M) \times f_{u,inc}) + C_{lumen,i} (\mu M)]} \times REF,$$

where  $P_{app,trans,n,i}$  represents apparent permeability contributed by the  $n^{th}$  transporter in the  $i^{th}$  segment of the intestine,  $A$  represents the transwell surface area,  $f_{u,inc}$  represents the fraction unbound in the incubation system,  $C_{lumen,i}$  represents the concentration of substrate in the lumen of the  $i^{th}$  segment.

From experiments conducted on Caco-2 cells (Ölander et al., 2016):

$$J_{max} = 255 \pm 65 \text{ pmol/min/mg protein}$$

$$K_m = 25 \mu M$$

$$\text{Insert growth area (A)} = 0.33 \text{ cm}^2$$

$$f_{u,inc} = 1$$

$$REF = 0.0625$$

To use this kinetic data in Simcyp, units of  $J_{max}$  must be converted to pmol/min. In a 12-well plate ( $A = 1.12 \text{ cm}^2$ ), a single filter contains  $0.346 \pm 0.04 \text{ mg}$  (mean  $\pm$ SD,  $n = 96$ ) of protein<sup>a</sup>.

Therefore,

$$J_{max} = 255 \pm 65 \text{ pmol/min/mg protein} \times 0.346 \text{ mg}/1.12 \text{ cm}^2 = 78.777 \text{ pmol/min/cm}^2$$

Since  $A = 0.33 \text{ cm}^2$  was used in our model, the following represents the input for  $J_{max}$ :

$$J_{max} = 78.777 \text{ pmol/min/cm}^2 \times 0.33 = 26.0 \text{ pmol/min}$$

<sup>a</sup> Personal communication with Magnus Ölander, 2018

### Results S4: Derivation of Intersystem Extrapolation Factors (ISEF) for Metabolism

1. The total  $CL_{int}$  due to metabolism of fluvastatin in HLM (Watanabe et al., 2010):

$$\begin{aligned} CL_{int,u}(HLM) &= J_{max} \div (K_m \times f_{u,inc}) \\ &= 5.57 \text{ ml/min/g liver} \div (48.8 \text{ mg protein/g liver} \times 0.308) \\ &= 370.6 \mu\text{L/min/mg protein} \end{aligned}$$

2. Using equation (15) from the methods section, ISEF for CYP3A4, 2C9 and 2C8 (Fischer et al., 1999):

$$\begin{aligned} ISEF &= \frac{CL_{int,u}(HLM) (\mu\text{L/min/mg protein})}{\sum_{j=1}^n \left[ \sum_{i=1}^n (CL_{int,i,u}(rhCYP_j) (\mu\text{L/min/pmol CYP}) \times CYP_j \text{ abundance}_{HLM} (\text{pmol CYP}_j/\text{mg protein})) \right]} \\ &= 370.6 \div \left[ \left( \frac{2 \text{ pmol/min/pmol CYP3A4}}{7.1 \mu\text{M} \times 0.308} \times 108 \text{ pmol/mg protein} \right) + \right. \\ &\quad \left. \left( \frac{0.078 \text{ pmol/min/pmol CYP2C9}}{0.9 \mu\text{M} \times 0.308} \times 93 \text{ pmol/mg protein} \right) + \right. \\ &\quad \left. \left( \frac{0.038 \text{ pmol/min/pmol CYP2C9}}{1.0 \mu\text{M} \times 0.308} \times 93 \text{ pmol/mg protein} \right) + \right. \\ &\quad \left. \left( \frac{0.038 \text{ pmol/min/pmol CYP2C9}}{1.8 \mu\text{M} \times 0.308} \times 93 \text{ pmol/mg protein} \right) + \right. \\ &\quad \left. \left( \frac{0.132 \text{ pmol/min/pmol CYP2C8}}{2.8 \mu\text{M} \times 0.308} \times 56 \text{ pmol/mg protein} \right) + \right] \\ &= 370.6 \div 151.363 \\ &= 2.444 \end{aligned}$$

## Results S5: Derivation of Inputs into Mech KiM for Rosuvastatin

All data is taken from Verhulst et al. (2008)

1. Number of proximal and distal tubular cells (PTC, DTC) per surface area of filter

$$= \frac{\text{Total number of cells seeded}}{\text{Surface area of the filter}} = \frac{50000 \text{ cells}}{0.33 \text{ cm}^2} = 0.1515 \times 10^6 \text{ cells/cm}^2$$

2. Clearance due to passive diffusion across DTC:

$$\begin{aligned} CL_{PD} &= \text{Rate of entry of rosuvastatin into DTC} \div \text{Concentration of drug in the medium} \\ &= 460.70 \text{ pmol/cm}^2/\text{h} \div 10 \text{ } \mu\text{mol/L} \\ &= 46.07 \text{ } \mu\text{l/cm}^2/\text{h} \div 0.1515 \times 10^6 \text{ cells/cm}^2 \div 60 \text{ min/h} \\ &= 0.00507 \text{ ml/min}/10^6 \text{ cells} \end{aligned}$$

3. Converting  $J_{max}$  to a function of  $10^6$  PTC:

$$J_{max} = 4951 \text{ pmol/cm}^2/\text{h} \div 0.1515 \times 10^6 \text{ cells/cm}^2 \div 60 \text{ min/h} = 546 \text{ pmol/min}/10^6 \text{ PTC}$$

**Tab. S1: Rosuvastatin input parameters**

Parameter	Value	Reference	Comments
Molecular weight (g/mol)	481.54	Jamei et al., 2014	
$\log P_{o:w}$	2.4	Jamei et al., 2014	
Compound type	Monoprotic acid	Jamei et al., 2014	
$pK_a$	4.27	Jamei et al., 2014	
Blood/plasma ratio	0.625	Jamei et al., 2014	
Fraction unbound in plasma	0.107	Jamei et al., 2014	
<b>Absorption</b>			
Absorption model	ADAM		
$f_{u_{gut}}$	1		Default value
$P_{eff,man}$ ( $10^{-4}$ cm/s)	0.1843941		Predicted in Simcyp
Permeability assay	Caco-2	Li et al., 2011	
Apical pH : basolateral pH	7.4 : 7.4	Li et al., 2011	
Activity	Passive & active	Li et al., 2011	
Apparent permeability ( $10^{-6}$ cm/s)	0.4	Li et al., 2011	Digitized plot
Reference compound	Propranolol		
Reference compound value ( $10^{-6}$ cm/s)	43		Default value in Simcyp
Scalar	1		
Formulation	Solution or tablet	Farshi, 2011; Potur, 2014	Refer to Table S5 and S6
<b>Distribution</b>			
Distribution model	Full PBPK model		
Tissue: plasma coefficient	Modified based on rat distribution data	Nezasa et al., 2002	Refer to Table S14
$V_{ss}$ (L/kg)	0.7002882	Rodgers and Rowland, 2007	Predicted in Simcyp
<b>Elimination</b>			
<b>Metabolism</b>			
Enzyme	CYP3A4		
Metabolic assay	HLM	Fujino et al., 2004b	
$CL_{int}$ ( $\mu$ l/min/mg protein)	1.1	Fujino et al., 2004b	
$f_{u_{mic}}$	0.937		Predicted in Simcyp
Enzyme	UGT 1A1		
rUGT system	BD Supersomes		
$V_{max}$ (pmol/min/mg protein)	17	Schirris et al., 2015	
$K_m$ ( $\mu$ M)	16	Schirris et al., 2015	
rUGT scalar	0.92	Simcyp Database	
Enzyme	UGT 1A3		
rUGT system	BD Supersomes		
$V_{max}$ (pmol/min/mg protein)	105	Schirris et al., 2015	
$K_m$ ( $\mu$ M)	220	Schirris et al., 2015	
rUGT scalar	1		Simcyp default value
<b>Hepatic Transport</b>			
$CL_{PD}$ (ml/min/million hepatocytes)	0.0025	Jamei et al., 2014	
$f_{u_{IW}}$	0.9673012		Predicted in Simcyp
$f_{u_{EW}}$	0.1869325		Predicted in Simcyp
Transporter	NTCP	Bi et al., 2013	

Transporter assay	Suspension hepatocyte		Difference in uptake with and without sodium
$CL_{int,T}$ ( $\mu\text{l}/\text{min}/\text{million}$ )	3.4	Bi et al., 2013	
REF	1.353	Bi et al., 2013; Vildhede et al., 2015	Refer to Table S11
Transporter	OATP1B1		
Transporter assay	HEK-293 cells	Izumi et al., 2018	Gene transfected cell
$J_{max}$ (pmol/min/million)	103	Izumi et al., 2018	
$K_m$ ( $\mu\text{M}$ )	9.31	Izumi et al., 2018	
REF	8.656	Vildhede et al., 2015; Izumi et al., 2018	Refer to Table S11
Transporter	OATP1B3		
Transporter assay	HEK-293 cells	Bosgra et al., 2014	Gene transfected cell
$J_{max}$ (pmol/min/million)	111.3	Bosgra et al., 2014	Refer to Results S2
$K_m$ ( $\mu\text{M}$ )	16.5	Bosgra et al., 2014	
REF	8.036	Bosgra et al., 2014; Vildhede et al., 2015	Refer to Table S11
Transporter	OATP2B1		
Transporter assay	HEK-293 cells	Bosgra et al., 2014	Gene transfected cell
$J_{max}$ (pmol/min/million)	17.3	Bosgra et al., 2014	Refer to Results S2
$K_m$ ( $\mu\text{M}$ )	26.1	Bosgra et al., 2014	
REF	100	Bosgra et al., 2014; Vildhede et al., 2015	Refer to Table S11
Transporter	MRP4		
Transporter assay	Membrane vesicles	Pfeifer et al., 2013	
$J_{max}$ (pmol/min/million)	1140	Pfeifer et al., 2013	
$K_m$ ( $\mu\text{M}$ )	21	Pfeifer et al., 2013	
REF	0.028	Pfeifer et al., 2013; Vildhede et al., 2015	Refer to Table S11
Transporter	Canalicular efflux		
Transporter assay	SCHH		
$CL_{int,T}$ ( $\mu\text{l}/\text{min}/\text{million}$ )	1.5	Jones et al., 2012	
REF	1.611	Vildhede et al., 2015	Refer to Table S11
<b>Renal Transporters</b>			
$CL_{PD,basal}$ (ml/min/million PTC)	0.00507	Verhulst et al., 2008	Digitized and calculated Refer to Results S5
$CL_{PD,apical}$ (ml/min/million PTC)	0.00507	Verhulst et al., 2008	Assumed to be equal to $CL_{PD,apical}$
$f_{uKidney,cell}$	0.985129		Predicted in Simcyp
$f_{uUrine}$	1		Default value in Simcyp
Transporter	OAT3		
Function	Uptake		
Transporter assay	PTC and DTC mixed monolayer	Verhulst et al., 2008	
$J_{max}$ (pmol/min/million cells)	546	Verhulst et al., 2008	Refer to Results S5
$K_m$ ( $\mu\text{M}$ )	20.4	Verhulst et al., 2008	

Transporter	MRP4		
Function	Efflux		
$J_{max}$ (pmol/min/million cells)	546		Assumed to be equal to uptake kinetics
$K_m$ ( $\mu$ M)	20.4		

Caco-2, colorectal adenocarcinoma cells;  $CL_{int}$ , intrinsic clearance;  $CL_{PD}$ , passive diffusion clearance;  $CL_{PD,apical}$ , passive diffusion clearance for the kidney between the renal cell and the tubule;  $CL_{PD,basal}$ , passive diffusion clearance for the kidney between the blood and renal cell; DTC, distal tubular cells;  $fU_{gut}$ , fraction unbound within the enterocyte;  $fU_{mic}$ , fraction unbound in microsomal system; HEK-293, human embryonic kidney cells; HLM, human liver microsome;  $J_{max}$ , maximum flux;  $K_m$ , Michaelis–Menten constant;  $K_p$ , partition coefficient; MRP4, multidrug resistance protein 4; NTCP, sodium-taurocholate co-transporting polypeptide; OAT3, organic anion transporter 3; OATP, organic anion-transporting polypeptide;  $P_{eff,man}$ , Effective human jejunum permeability;  $pK_a$ , acid dissociation constant; PTC, proximal tubular cells; REF, relative expression factor; SCHH, sandwich-cultured human hepatocyte;  $V_{ss}$ , apparent volume of distribution at steady state

**Tab. S2: Fluvastatin input parameters**

Parameter	Value	Reference	Comments
Molecular weight (g/mol)	411.5	Winiwarter et al., 1998	
logP <sub>o:w</sub>	4.17	Winiwarter et al., 1998	
Compound type	Monoprotic acid	Winiwarter et al., 1998	
pK <sub>a</sub>	4.31	Winiwarter et al., 1998	
Blood/plasma ratio	0.55	Tse et al., 1993	
Fraction unbound in plasma	0.009	Tse et al., 1993	
<b>Absorption</b>			
Absorption model	ADAM		
<i>f</i> <sub>gut</sub>	1		Default value
<i>P</i> <sub>eff,man</sub> (10 <sup>-4</sup> cm/s)	3.069295		Predicted in Simcyp
Permeability assay	Caco-2	Lindahl et al., 2004	
Apical pH : basolateral pH	6.5 : 7.4	Lindahl et al., 2004	
Activity	Passive & active	Lindahl et al., 2004	
Apparent permeability (10 <sup>-6</sup> cm/s)	16.237	Lindahl et al., 2004	Digitized data
Reference compound	Verapamil		
Reference compound value (10 <sup>-6</sup> cm/s)	20.6		Default value in Simcyp
Scalar	1		
Formulation	Solution		
<b>Distribution</b>			
Distribution model	Full PBPK model		
V <sub>ss</sub> (l/kg)	0.09742813	Rodgers and Rowland, 2007	Predicted in Simcyp
<b>Elimination</b>			
<b>Metabolism</b>			
Enzyme	CYP3A4		
Metabolism assay	rhCYP	Fischer et al., 1999	
Metabolic pathway	Pathway 2	Fischer et al., 1999	5-hydroxylation
<i>V</i> <sub>max</sub> (pmol/min/pmol)	2	Fischer et al., 1999	
<i>K</i> <sub>m</sub> (μM)	7.1	Fischer et al., 1999	
<i>f</i> <sub>mic</sub>	0.308	Watanabe et al., 2010	
ISEF	2.444	Watanabe et al., 2010; Nakamura et al., 2016	Refer to Results S4
Enzyme	CYP2C9		
Metabolism assay	rhCYP	Fischer et al., 1999	
Metabolic pathway	Pathway 1	Fischer et al., 1999	6-hydroxylation
<i>V</i> <sub>max</sub> (pmol/min/pmol)	0.0783	Fischer et al., 1999	
<i>K</i> <sub>m</sub> (μM)	0.9	Fischer et al., 1999	
<i>f</i> <sub>mic</sub>	0.308	Watanabe et al., 2010	
ISEF	2.444	Watanabe et al., 2010; Nakamura et al., 2016	Refer to Results S4
Enzyme	CYP2C9		
Metabolism assay	rhCYP	Fischer et al., 1999	
Metabolic pathway	Pathway 2	Fischer et al., 1999	5-hydroxylation
<i>V</i> <sub>max</sub> (pmol/min/pmol)	0.0383	Fischer et al., 1999	
<i>K</i> <sub>m</sub> (μM)	1	Fischer et al., 1999	
<i>f</i> <sub>mic</sub>	0.308	Watanabe et al., 2010	
ISEF	2.444	Watanabe et al., 2010; Nakamura et al., 2016	Refer to Results S4

Enzyme	CYP2C9		
Metabolism assay	rhCYP	Fischer et al., 1999	
Metabolic pathway	Pathway 3	Fischer et al., 1999	N-deisopropyl
$V_{max}$ (pmol/min/pmol)	0.0383	Fischer et al., 1999	
$K_m$ ( $\mu$ M)	1.8	Fischer et al., 1999	
$f_{u_{mic}}$	0.308	Watanabe et al., 2010	
ISEF	2.444	Watanabe et al., 2010; Nakamura et al., 2016	Refer to Results S4
Enzyme	CYP2C8		
Metabolism assay	rhCYP	Fischer et al., 1999	
Metabolic pathway	Pathway 2	Fischer et al., 1999	5-hydroxylation
$V_{max}$ (pmol/min/pmol)	0.1317	Fischer et al., 1999	
$K_m$ ( $\mu$ M)	2.8	Fischer et al., 1999	
$f_{u_{mic}}$	0.308	Watanabe et al., 2010	
ISEF	2.444	Watanabe et al., 2010; Nakamura et al., 2016	Refer to Results S4
$CL_R$ (l/h)	0		Default value in Simcyp
<b>Hepatic transport</b>			
$CL_{PD}$ (ml/min/million hepatocytes)	0.025	Bi et al., 2013	
$f_{u_{IW}}$	0.3146003		Predicted in Simcyp
$f_{u_{EW}}$	0.0171335		Predicted in Simcyp
Transporter	Sinusoidal uptake		
Transporter assay	SCHH		
$CL_{int,T}$ ( $\mu$ l/min/million hepatocytes)	158.1	Izumi et al., 2018	
REF	3.55		Refer to Table S12
Transporter	Canalicular efflux		
Transporter assay	SCHH		
$CL_{int,T}$ ( $\mu$ l/min/million hepatocytes)	17	Jones et al., 2012	
RAF/REF	1.627		Refer to Table S12

$CL_{int}$ , intrinsic hepatic clearance;  $CL_{PD}$ , passive diffusion clearance;  $CL_R$ , renal clearance;  $f_{u_{gut}}$ , fraction unbound within the enterocyte;  $f_{u_{mic}}$ , fraction unbound in microsomal system;  $J_{max}$ , maximum flux;  $K_m$ , Michaelis–Menten constant;  $K_p$ , partition coefficient;  $P_{eff,man}$ , Effective human jejunum permeability;  $pK_a$ , acid dissociation constant; PTC, proximal tubular cells; REF, relative expression factor; rhCYP, recombinant human CYP450; SCHH, sandwich-cultured human hepatocyte;  $V_{ss}$ , apparent volume of distribution at steady state

**Tab. S3: Pitavastatin input parameters**

Parameter	Value	Reference	Comment
Molecular weight (g/mol)	421.49	FDA, 2012	
logP <sub>o,w</sub>	1.49	Kajinami et al., 2003	
Compound type	Monoprotic acid		
pK <sub>a</sub>	5.31	FDA, 2012	
Blood:plasma ratio	0.55	FDA, 2012	
Fraction unbound in plasma	0.0052	Watanabe et al., 2011	
<b>Absorption</b>			
Absorption model	ADAM model		
$f_{ugut}$	1		Default value in Simcyp
$P_{eff,man}$ (10 <sup>-4</sup> cm/s)	1.023977		Predicted in Simcyp
Permeability assay	LLC-PK1	Shirasaka et al., 2011	
Value	4.16	Shirasaka et al., 2011	
Reference compound	Propranolol		
Reference compound value (10 <sup>-6</sup> cm/s)	36		Default value in Simcyp
Scalar	1		
Formulation	Solution		Assumption due to lack of dissolution data
<b>Intestinal Transporter</b>			
Transporter	OATP2B1		
$J_{max}$ (pmol/min/cm <sup>2</sup> )	78.777	Ölander et al., 2016	Refer to Results S3
$K_m$ (μM)	25	Ölander et al., 2016	
$f_{uinc}$	1		
Insert growth area of the Transwell (cm <sup>2</sup> )	0.33		
System	User		
RAF/REF	0.0625	Ölander et al., 2016	
<b>Distribution</b>			
Distribution model	Full PBPK model		
Tissue:plasma coefficient	Modified based on rat distribution data	Kimata et al., 1998	Table S15 and S16
V <sub>ss</sub> (L/kg)	0.5103121	Rodgers and Rowland, 2007	Predicted in Simcyp
<b>Elimination</b>			
Enzyme	CYP2C9		
Metabolic assay	HLM	Fujino et al., 2004b	Allocated to CYP3A4
$CL_{int}$ (μl/min/mg protein)	2.5	Fujino et al., 2004b	
$f_{umic}$	0.962		Predicted in Simcyp
Enzyme	UGT 1A3		
Metabolic assay	BD Supersomes	Schirris et al., 2015	

$V_{max}$ (pmol/min/mg protein)	880	Schirris et al., 2015	
$K_m$ ( $\mu$ M)	10	Schirris et al., 2015	
rUGT scaler	1.0		Simcyp default value
Enzyme	UGT 2B7		
Metabolic assay	BD Supersomes	Schirris et al., 2015	
$V_{max}$ (pmol/min/mg protein)	12900	Schirris et al., 2015	
$K_m$ ( $\mu$ M)	220	Schirris et al., 2015	
rUGT scalar	2.81	Simcyp Database	
<b>Hepatic Transporters</b>			
$CL_{PD}$ (ml/min/million hepatocytes)	0.00773	Fujino et al., 2004a	
$f_{u_{IW}}$	0.9566636		Predicted in Simcyp
$f_{u_{EW}}$	0.009929368		Predicted in Simcyp
Transporter	NTCP		
Transporter assay	Suspension hepatocyte	Bi et al., 2013	Difference in uptake with and without sodium
$CL_{int,T}$ ( $\mu$ l/min/million)	20	Bi et al., 2013	
RAF/REF	1.353	Bi et al., 2013; Vildhede et al., 2015	Refer to Table S13
Transporter	OATP1B1		
Transporter assay	HEK-293 cells	Hirano, 2004	Gene transfected cells
$J_{max}$ (pmol/min/million)	230	Hirano, 2004	
$K_m$ ( $\mu$ M)	3	Hirano, 2004	
RAF/REF	5.936	Hirano, 2004; Vildhede et al., 2015	Refer to Table S13
Transporter	OATP1B3		
Transporter assay	HEK-293 cells	Hirano, 2004	Gene transfected cells
$J_{max}$ (pmol/min/million)	100	Hirano, 2004	
$K_m$ ( $\mu$ M)	3.25	Hirano, 2004	
RAF/REF	4.904	Hirano, 2004; Vildhede et al., 2015	Refer to Table S13
Transporter	OATP2B1		
Transporter assay	HEK-293 cells	Hirano et al., 2006	Gene transfected cells
$J_{max}$ (pmol/min/million)	7.36	Hirano et al., 2006	
$K_m$ ( $\mu$ M)	1.17	Hirano et al., 2006	
RAF/REF	0.19	Hirano et al., 2006; Vildhede et al., 2015	Refer to Table S13
Transporter	MRP3		
Transporter assay	Membrane vesicles	Vildhede et al., 2016	
$J_{max}$ (pmol/min/million)	2380	Vildhede et al., 2016	
$K_m$ ( $\mu$ M)	448	Vildhede et al., 2016	
RAF/REF	0.025	Vildhede et al., 2015, 2016	Refer to Table S13

Transporter	P-gp/MDR1		
Transporter assay	Membrane vesicles	Vildhede et al., 2016	
$J_{max}$ (pmol/min/million)	433	Vildhede et al., 2016	
$K_m$ ( $\mu$ M)	83.7	Vildhede et al., 2016	
RAF/REF	0.00181	Vildhede et al., 2015, 2016	Refer to Table S13
Transporter	BCRP		
Transporter assay	Membrane vesicles	Vildhede et al., 2016	
$J_{max}$ (pmol/min/million)	95.3	Vildhede et al., 2016	
$K_m$ ( $\mu$ M)	1.2	Vildhede et al., 2016	
RAF/REF	0.00052	Vildhede et al., 2015, 2016	Refer to Table S13
<b>Renal Transporters</b>			
$CL_{PD,basal}$ (ml/min/PTC)	0.0151	Watanabe et al., 2011	Calculated. Refer to Results S1
$CL_{PD,apical}$ (ml/min/million PTC)	0.0151		Assumed to be equal to $CL_{PD,basal}$
$f_{u_{Kidney,cell}}$	0.9800262		Predicted in Simcyp
$f_{u_{Urine}}$	1		Default value in Simcyp
Transporter	OAT3		
Transporter Assay	Human kidney slices		
Function	Uptake		
$J_{max}$ (pmol/min/million cells)	12.672	Watanabe et al., 2011	Refer to Results S1
$K_m$ ( $\mu$ M)	3.3	Fujino et al., 2005	BD Supersomes
Transporter	MRP4		
Function	Efflux		
$J_{max}$ (pmol/min/million cells)	12.672		Assumed to be equal to uptake kinetics
$K_m$ ( $\mu$ M)	3.3		

BCRP, breast cancer resistant protein; Caco-2, colorectal adenocarcinoma cells;  $CL_{int}$ , intrinsic hepatic clearance;  $CL_{PD}$ , passive diffusion clearance;  $CL_{PD,apical}$ , passive diffusion clearance for the kidney between the renal cell and the tubule;  $CL_{PD,basal}$ , passive diffusion clearance for the kidney between the blood and renal cell; DTC, distal tubular cells;  $f_{u_{gut}}$ , fraction unbound within the enterocyte;  $f_{u_{mic}}$ , fraction unbound in microsomal system; HEK-293, human embryonic kidney cells; HLM, human liver microsomes;  $J_{max}$ , maximum flux;  $K_m$ , Michaelis-Menten constant;  $K_p$ , partition coefficient; MRP4, multidrug resistance protein 4; NTCP, sodium-taurocholate co-transporting polypeptide; OAT3, organic anion transporter 3; OATP, organic anion-transporting polypeptide;  $P_{eff,man}$ , Effective human jejunum permeability; P-gp, p-glycoprotein;  $pK_a$ , acid dissociation constant; PTC, proximal tubular cells; REF, relative expression factor; SCHH, sandwich-cultured human hepatocyte;  $V_{ss}$ , apparent volume of distribution at steady state

**Tab. S4: Referenced clinical data**

Drug	Route of Administration	Dose (mg)	Dosing Frequency	Number of participants	Proportion of females	Reference
Rosuvastatin	IV infusion	8	Over 4 hours	10	0	Martin et al., 2003
	Oral	20	Single dose	7	0.43	Wu et al., 2017
	Oral	10	Once daily for 14 days	21	0.083	Martin et al., 2002
Fluvastatin	IV infusion	2	Over 20 minutes	6	0	Tse et al., 1992
	Oral	10	Single dose	6	0	Tse et al., 1992
	Oral	40	Once daily for 6 days	6	0	Tse et al., 1992
Pitavastatin	IV	2	Over 1 hour	18	0	FDA, 2012
	Oral	2	Single dose	18	0	FDA, 2012
	Oral	4	Once daily for 5 days	18	0	FDA, 2012

**Tab. S5: Dissolution rate for rosuvastatin 20 mg**

At pH = 1.2				
Time (h)	0.25	0.5	0.75	1
% Dissolved	52.3	56.85	57.5	57.5
At pH = 6.8				
Time (h)	0.25	0.5	0.75	1
% Dissolved	96.45	96.95	97.32	97.39

Dissolution data of Crestor<sup>®</sup> 20 mg in pH 6.8 and 1.2 were used (Potur, 2014).

**Tab. S6: Dissolution rate for rosuvastatin 10 mg**

At pH = 1.2							
Time (h)	0.0833	0.167	0.25	0.333	0.5	0.75	1
% Dissolved	18.1	41.6	55.1	64.1	75.9	85.2	89.8
At pH = 6.8							
Time (h)	0.25	0.5	0.75	1			
% Dissolved	100	100	100	100			

Dissolution data of Crestor<sup>®</sup> 10 mg (pH = 8.6) and 40 mg (pH = 1.2) were used (Farshi, 2011; Potur, 2014).

**Tab. S7: Derivation of SF1 values for rosuvastatin**

Hepatic Transporter	<i>In vitro</i> system		Isolated hepatocyte system		SF1
	Type of system	Activity/expression	Type of system	Activity/expression	
NTCP	-	-	Hepatocyte suspension	3.4 <sup>a</sup> μl/min/10 <sup>6</sup> cells	-
OATP1B1	HEK-293	127 <sup>b</sup> μl/min/mg protein	Hepatocyte suspension	332 <sup>b</sup> μl/min/10 <sup>6</sup> cells	2.610 mg protein/10 <sup>6</sup> cells
OATP1B3	HEK-293	1.4 <sup>c</sup> pmol/10 <sup>6</sup> cells	Primary hepatocyte	2.2 <sup>c</sup> pmol/10 <sup>6</sup> cells	1.571
OATP2B1	HEK-293	0.2 <sup>c</sup> pmol/10 <sup>6</sup> cells	Primary hepatocyte	24.2 <sup>c</sup> pmol/10 <sup>6</sup> cells	121
MRP4	Membrane vesicles	54.3 <sup>d</sup> μl/min/mg protein	SCHH	0.85 <sup>d,e</sup> μl/min/10 <sup>6</sup> cells	0.0157 mg protein/10 <sup>6</sup> cells
Canalicular efflux	-	-	SCHH	17 <sup>f</sup> μl/min/10 <sup>6</sup> cells	-

<sup>a</sup>Bi et al. (2013); <sup>b</sup>Izumi et al. (2018); <sup>c</sup>Bosgra et al. (2014); <sup>d</sup>Pfeifer et al. (2013); <sup>e</sup>Basolateral efflux clearance in SCHH was scaled by the number of hepatocyte per gram of liver obtained from Simcyp (118 x 10<sup>6</sup> HHEP/g); <sup>f</sup>Jones et al. (2012)

**Tab. S8: Derivation of SF1 values for pitavastatin**

Hepatic transporter	<i>In vitro</i> system		Isolated hepatocyte system		SF1
	Type of system	Activity/expression	Type of system	Activity/expression	
NTCP	-	-	Hepatocyte suspension	20 <sup>a</sup> $\mu$ l/min/10 <sup>6</sup> cells	-
OATP1B1	HEK-293	-	Primary hepatocyte	-	1.790 <sup>b</sup> mg protein/10 <sup>6</sup> cells
OATP1B3	HEK-293	-	Primary hepatocyte	-	0.959 <sup>b</sup> mg protein/10 <sup>6</sup> cells
OATP2B1	HEK-293	-	Primary hepatocyte	-	0.155 <sup>b</sup> mg protein/10 <sup>6</sup> cells
MRP3	Membrane vesicles	40.0 <sup>c</sup> pmol/mg vesicular protein	SCHH	0.417 <sup>c,d</sup> pmol/10 <sup>6</sup> cells	0.0105 mg protein/10 <sup>6</sup> cells
BCRP	Membrane vesicles	260 <sup>c</sup> pmol/mg vesicular protein	SCHH	0.0841 <sup>c,d</sup> pmol/10 <sup>6</sup> cells	0.000327 mg protein/10 <sup>6</sup> cells
P-gp	Membrane vesicles	280 <sup>c</sup> pmol/mg vesicular protein	SCHH	0.3167 <sup>c,d</sup> pmol/10 <sup>6</sup> cells	0.00113 mg protein/10 <sup>6</sup> cells

<sup>a</sup>Bi et al. (2013); <sup>b</sup>Hirano (2004); <sup>c</sup>Vildhede et al. (2016); <sup>d</sup>Expression of transporters in SCHH was scaled by 0.3235 mg protein/10<sup>6</sup> HHEP obtained by dividing amount of protein per well (0.1214 mg) with number of HHEP per well (0.375 x 10<sup>6</sup> HHEP)

**Tab. S9: Derivation of SF2 from uptake transporters**

Hepatic uptake transporter	Absolute abundance in (pmol/mg membrane protein)		Ratio of abundance in liver to hepatocyte, SF2
	Liver membrane	Hepatocyte membrane	
OATP1B1	19.600	5.910	3.316
OATP1B3	2.700	0.528	5.114
NTCP	1.880	0.414	4.541
OATP2B1	1.550	1.260	1.230

Values are obtained from Vildhede et al. unless otherwise stated (Vildhede et al., 2015).

**Tab. S10: Derivation of SF2 from efflux transporters**

Hepatic efflux transporter	Absolute abundance in (pmol/mg membrane protein)		Ratio of abundance in liver to hepatocyte, SF2
	Liver membrane	Hepatocyte membrane	
BSEP	3.050	1.580	1.930
MRP6	1.370	0.844	1.623
MRP2	0.815	0.501	1.627
P-gp	0.576	0.360	1.600
MRP3	0.407	0.171	2.380
BCRP	0.419 <sup>a</sup>	0.263 <sup>b</sup>	1.593
Average	1.106	0.646	1.793

Values are obtained from Vildhede et al. unless otherwise stated.

<sup>a</sup>Ohtsuki et al. (2011); <sup>b</sup>Vildhede et al. (2016)

**Tab. S11: Derivation of REF for rosuvastatin**

Hepatic transporter	Scaling factor 1 (SF1)	Scaling factor 2 (SF2)	REF
NTCP	-	1.353 <sup>a</sup>	1.353
OATP1B1	2.610	3.316	8.656
OATP1B3	1.571	5.114	8.036
OATP2B1	121	1.230	148.830
MRP4	0.0157	1.793 <sup>b</sup>	0.028
Canalicular efflux	-	1.610 <sup>c</sup>	1.610

SF1 and SF2 are obtained from Table S7 – S10 unless otherwise stated.

<sup>a</sup> Obtained from transporter expression in Bi et al. (2013) and Vildhede et al. (2015); <sup>b</sup> Average value in Table S10 used; <sup>c</sup> Average value of BCRP and MRP2 ratio in Table S10 used

**Tab. S12: Derivation of REF for fluvastatin**

Hepatic transporter	Scaling factor 1 (SF1)	Scaling factor 2 (SF2)	REF
Sinusoidal uptake	-	3.55 <sup>a</sup>	3.55
Canalicular efflux	-	1.627 <sup>b</sup>	1.627

<sup>a</sup> Average of OATP1B1, OATP1B3, OATP2B1 and NTCP from Table S9; <sup>b</sup> Values for MRP2 in Table S10

**Tab. S13: Derivation of REF for pitavastatin**

Hepatic transporter	Scaling factor 1 (SF1)	Scaling factor 2 (SF2)	REF
NTCP	-	1.353 <sup>a</sup>	1.353
OATP1B1	1.790	3.316	5.936
OATP1B3	0.959	5.114	4.904
OATP2B1	0.155	1.230	0.190
MRP3	0.0105	2.380	0.0249
P-gp	0.00113	1.600	0.00181
BCRP	0.000327	1.593	0.00052

SF1 and SF2 are obtained from Table S7-S10 unless otherwise stated.

<sup>a</sup> Obtained from transporter expression in Bi et al. (2013) and Vildhede et al. (2015)

**Tab. S14: Tissue concentration of radioactivity and tissue-to-plasma equilibrium distribution ratio ( $K_p$ ) for rosuvastatin**

Tissue concentration of radioactivity (ng.eq/g or ng.eq/ml) and  $K_p$  after single oral administration of <sup>14</sup>C-rosuvastatin at 5 mg/kg in male Sprague-Dawley rats at various time points (Nezasa et al., 2002).

Organs	Time at									
	15 minute		1.5 hours		4 hours		8 hours		24 hours	
	Radioactivity	$K_p$	Radioactivity	$K_p$	Radioactivity	$K_p$	Radioactivity	$K_p$	Radioactivity	$K_p$
Plasma	38.600	1.000	79.400	1.000	68.100	1.000	44.800	1.000	28.900	1.000
Heart	17.500	<b>0.453</b>	31.500	0.397	27.700	0.407	16.600	0.371	9.000	0.311
Lung	16.500	0.427	31.700	0.399	31.600	<b>0.464</b>	16.700	0.373	n.d.	-
Spleen	11.300	<b>0.293</b>	18.000	0.227	14.400	0.211	8.800	0.196	7.000	0.242
Pancreas	26.100	<b>0.676</b>	34.300	0.432	26.400	0.388	18.500	0.413	n.d.	-
Fat	n.d.	-	41.500	0.523	69.500	<b>1.021</b>	27.700	0.618	15.700	0.543
Brown fat	33.700	0.873	65.500	0.825	57.200	0.840	30.000	0.670	25.300	<b>0.875</b>
Skeletal muscle	n.d.	-	15.800	0.199	14.100	<b>0.207</b>	n.d.	-	n.d.	-
Skin	11.800	0.306	30.100	0.379	34.800	0.511	23.000	<b>0.513</b>	10.000	0.346

n.d., not detected

Values in bold represent the largest  $K_p$  value observed and are used in our model.

**Tab. S15: Tissue concentration of unchanged drug and  $K_p$  for unchanged pitavastatin**

Concentration of unchanged pitavastatin ( $\mu\text{g/g}$ ) and  $K_p$  after single oral administration of  $^{14}\text{C}$ -pitavastatin at 1 mg/kg in male Sprague-Dawley rats at various time points (Kimata et al., 1998).

Organs	Time at							
	0.5 hour		1 hour		3 hours		6 hours	
	Concentration	$K_p$	Concentration	$K_p$	Concentration	$K_p$	Concentration	$K_p$
Plasma	0.2	1	0.079	1	0.033	1	0.013	1
Heart	0.101	0.505	0.041	<b>0.519</b>	0.007	0.212	n.d.	-
Lung	0.131	0.655	0.034	0.430	0.017	0.515	0.011	<b>0.846</b>
Skeletal muscle	0.019	0.095	0.011	<b>0.139</b>	0.003	0.091	n.d.	-

n.d., not detected

Values in bold represent the largest  $K_p$  value observed and are used in our model.

**Tab. S16: Tissue concentration of total radioactivity and  $K_p$  for pitavastatin**

Tissue concentration of radioactivity (ng.eq/g or ng.eq/ml) and  $K_p$  after single oral administration of  $^{14}\text{C}$ -pitavastatin at 1 mg/kg in male Sprague-Dawley rats at various time points (Kimata et al., 1998).

Organs	Time at							
	15 minute		1 hour		6 hours		24 hours	
	Radioactivity	$K_p$	Radioactivity	$K_p$	Radioactivity	$K_p$	Radioactivity	$K_p$
Plasma	0.143	1.000	0.103	1.000	0.04	1.000	0.012	1.000
Brain	0.009	<b>0.063</b>	0.006	0.058	n.d.	-	n.d.	-
Spleen	0.026	0.182	0.021	<b>0.204</b>	0.008	0.200	n.d.	-
Pancreas	0.051	0.357	0.044	0.427	0.02	<b>0.500</b>	n.d.	-
Abdominal fat	0.025	0.175	0.030	0.291	0.023	0.575	0.016	<b>1.333</b>
Skin	0.015	0.105	0.020	0.194	0.008	0.200	0.004	<b>0.333</b>
Bone marrow	0.032	<b>0.224</b>	0.022	0.214	0.007	0.175	n.d.	-

n.d., not detected

Values in bold represents the largest  $K_p$  value observed and are used in our model.

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